Recent updates on genomics and transcriptomics.

Isheng Jason Tsai

生圖教育訓練課程 2021.01.11







Lab setup

Molecular Biology & Sequencing R&D

NANOPORE

GridION

Analysis and algorithm development

Microbial Ecology & establishing collections

#tetrischallenge

Evolutionary genomics

million years ago

Comparative genomics/transcriptomics



Population genomics



Metagenomics/ metatranscriptomics



Tsai *et al* (2008, 2010) PNAS Liti *et al.*, Nature (2009) Tsai *et al.*, Nature (2013) Valentim *et al.*, Science (2013) Foth and Tsai *et al.*, Nature Genetics (2014) Hunt and Tsai *et al.*, Nature Genetics (2016) Natsumi and Tsai *et al.*, Nature Communications (2018) Coghlan *et al.*, Nature Genetics (2018) Chaw *et al.*, Nature Plants (2019) Sung *et al.*, CMGH (2019) Ke *et al.*, PNAS (2020) Lin *et al* Gut Microbes (2021)

Recent updates in...

Genomics

"is an interdisciplinary field of biology focusing on the structure, function, evolution, mapping, and editing of genomes." (wiki)

Systems Biology

"...is an approach in biomedical research to understanding the larger picture—be it at the level of the organism, tissue, or cell—by putting its pieces together."¹ (opposite to reductionist view of taking things apart)

Transcriptomics

"the study of the complete set of RNA transcripts that are produced by the genome, under specific circumstances or in a specific cell—using highthroughput methods, such as microarray analysis." (nature portfolio)

Bioinformatics

"is a subdiscipline of biology and computer science concerned with the acquisition, storage, analysis, and dissemination of biological data, most often DNA and amino acid sequences."

Lecture outline

- 1. Introduction
- 2. Brief history of sequencing
- 3. dawn of third gen sequencing
- 4. Case studies genomics
- 5. Case studies transcriptomics

This is how scientists see the world





基因 (gene): 一個有功能的DNA 片段 coding DNA: 可以轉譯成蛋白質的 DNA #noncoding 基因體 (genome): 物種一個細胞核內所有的DNA 定序 (sequencing): 解析出DNA 序列 [ATCGTGACGTGACGTAC...]

Genome



Genome = Parts list of a single genome

A typical genome/transcriptomic project



A 2021 genome/transcriptomic project (take home message)



More in-depth Analysis

Why sequence a genome?

- Phylogenetic position
- Differences between species (comparative genomics)
- Variations between individuals (population genetics)
- Help to understand biology
- Of economic, agricultural, medical, ecology values
- Help to understand biology

Things to consider in sequencing

- 1. Length
- 2. Depth
- 3. Biases
- 4. Errors

Read length matters in sequencing



Figure 5. Two copies of a repeat along a genome. The reads colored in red and those colored in yellow appear identical to the assembly program.



Figure 6. Genome mis-assembled due to a repeat. The assembly program incorrectly combined the reads from the two copies of the repeat leading to the creation of two separate contigs

https://www.cbcb.umd.edu/research/assembly_primer

Read length matters in sequencing



Depth matters in sequencing

10X

1X

ATCGATGACTGACTGAATGGTTGAC ATCGATGACTGACTGAATGGTTGAC ATCCATGACTGACTGAATGGTTGAC ATCGATGACTGACTGAATGGTTGAC ATCGATGACTGACTGAATGGTTGAC ATCGATGACTGAGTGAATGGTTGAC ATCGATGACTGAGTGAATGGTTGAC ATCGATGACTGAGTGAATGGTTGAC ATCGATGACTGAGTGAATGGTTGAC ATCGATGACTGAGTGAATGGTTGAC

Homozygous? Heterozygous? ATCGATCACTGACTGACTGGTTGAC

...ATCGATGACTGACTGACTGGTTGAC...

reference

Sequencing Biases

c Uniformity of sequence coverage according to GC content



http://www.nature.com/nrg/journal/v16/n11/fig_tab/nrg3933_F2.html

Sequencing Errors

A) Illustration of errors in Illumina data after a long homopolymer tract. Ion torrent data has a drop of coverage and multiple indels are visible in PacBio data.

B) Example of errors associated with short homopolymer tracts. Multiple insertions are visible in the PacBio Data... MiSeq sequences read generally correct through the homopolymer tract.



Quail et al., BMC Genomics (2012) 13:341

Sequencing – a brief history



Genomic sequencing

- 1. The Human Genome Project
- 2. Sequencing the unculturable majority
- 3. Sequencing the next generation
- 4. ChIP-seq captures the chromatin landscape
- 5. The dawn of personal genomes
- 6. A sequencing revolution in cancer
- 7. Transcriptomes a new layer of complexity
- 8. Long reads become a reality
- 9. Exploring whole exomes
- 10. Probing nuclear architecture with Hi-C
- 11. Sequencing one cell at a time
- 12. Waking the dead: sequencing archaic hominin genomes
- 13. Cataloguing a public genome
- 14. Our most elemental encyclopaedia
- 15. Pan-genomes: moving beyond the reference
- 16. Genomes go platinum
- 17. Filling in the gaps telomere to telomere

Proc. Natl. Acad. Sci. USA Vol. 74, No. 12, pp. 5463-5467, December 1977 Biochemistry

DNA sequencing with chain-terminating inhibitors

(DNA polymerase/nucleotide sequences/bacteriophage \$\$\phi_X174\$)

F. SANGER, S. NICKLEN, AND A. R. COULSON

Medical Research Council Laboratory of Molecular Biology, Cambridge CB2 2QH, England

Contributed by F. Sanger, October 3, 1977



http://www.ncbi.nlm.nih.gov/pmc/articles/PMC431765/

ABI 3730xi at TIGR (1.6Mb per day)



https://www.flickr.com/photos/jurvetson/57080968



Nature **409**, 860-921(15 February 2001) doi:10.1038/35057062



1442-2551 May 5567 Propert 1145-1414 (59

THE HUMAN GENOME

> AMERICAN ASSOCIATION TOP ADVANCEMENT OF SCHENCE



Calculating the economic impact of the Human Genome Project

Public funding of scientific R&D has a significant positive impact on the wider economy, but quantifying the exact impact of research can be difficult to assess. A new report by research firm Battelle Technology Partnership Practice estimates that **between 1988 and 2010, federal investment in genomic research generated an economic impact of \$796 billion**, which is impressive considering that Human Genome Project (HGP) spending **between 1990-2003 amounted to \$3.8 billion**. This figure equates to a return on investment (ROI) of 141:1 (that is, every \$1 invested by the U.S. government generated \$141 in economic activity). The report was commissioned by Life Technologies Foundation.

https://www.genome.gov/27544383/calculating-theeconomic-impact-of-the-human-genome-project/

2000-2010s – Second generation sequencing and associated challenges



https://www.nlm.nih.gov/about/https://www.nlm.nih.gov/about/2018CJ.html http://www.nature.com/news/2010/100331/full/464670a.html

A brief history of bioinformatics

Jeff Gauthier, Antony T Vincent, Steve J Charette, Nicolas Derome *Briefings in Bioinformatics* (2018) <u>https://doi.org/10.1093/bib/bby063</u> NGS = sequencing made cheaper, faster and higher throughput

World competing for sequencing power



INTRODUCTION

History of DNA sequencing – Main players' first commercial products and M&A



Clip slide

Illumina machines



Decreasing Price Per Gigabase (Gb)

Illumina: sequencing by synthesis

https://www.youtube.com/watch?v=fCd6B5HRaZ8



Illumina platform comparison

0.85

Cycles Throughput (Gb) Price Kit/Cartridge (GBP) Price per Gb (GBP) 100 200 300 120 100 200 -300 8:32 300-9.89 _300____500 10.71 200

NovaSeg 6000 S2 Reagent Kit (300 cycles) NovaSeg 6000 S1 Reagent Kit (300 cycles) NovaSeg 6000 S4 Reagent Kit (200 cycles) NovaSeq 6000 SP Reagent Kit (500 cycles) NovaSeg 6000 SP Reagent Kit (300 cycles) NovaSeg 6000 S2 Reagent Kit (200 cycles) NovaSeg 6000 S1 Reagent Kit (200 cycles) NovaSeg 6000 S2 Reagent Kit (100 cycles) HiSeq 3000/4000 SBS Kit (300 cycles) NovaSeq 6000 S1 Reagent Kit (100 cycles) NextSeg 2000 P3 NovaSeg 6000 SP Reagent Kit (100 cycles) HiSeg 3000/4000 SBS Kit (150 cycles) NextSeg 500/550 High Output Kit v2.5 (300 Cy... NextSeg 500/550 Mid Output Kit v2.5 (300 Cy ... NextSea 500/550 High Output Kit v2.5 (150 Cv... NextSeg 500/550 High Output Kit v2.5 (75 Cyc... NextSeq 500/550 Mid Output Kit v2.5 (150 Cy... HiSeg 3000/4000 SBS Kit (50 cycles) MiSeq Reagent Kit v3 (600-cycle) MiSeg Reagent Kit v2 MiniSeq High Output Reagent Kit (300-cycles) MiniSeg Mid Output Kit (300-cycles) MiniSeq High Output Reagent Kit (150-cycles) MiSeg Reagent Micro Kit v2 MiniSeg High Output Reagent Kit (75-cycles) iSeq 100 0.5 MiSeq Reagent Nano Kit v2 MiSeq Reagent Kit v2 MiSeg Reagent Nano KR. 32

NovaSeg 6000 S4 Reagent Kit (300 cycles)

NextSeq 2000 P2

NextSeg 2000 P2

NextSeq 2000 P2

NextSeq 2000 P3

NextSeg 2000 P3



And the arrival of 3rd generation sequencing... (much longer read lengths)

PacBio (Pacific Biosciences)





Sequel II

RSII

PacBio (Pacific Biosciences)

Half of data in reads: >190 kb Data per SMRT Cell: Up to 50 Gb



https://www.pacb.com/smrt-science/smrt-sequencing/smrt-sequencing-modes/
Oxford Nanopore



Oxford Nanopore – how it works

Introduction to nanopore https://vimeo.com/297106166

Voltrax https://vimeo.com/297106291

Sequencing for farmers https://vimeo.com/294216876

@ Oceans https://vimeo.com/294744892 Rainforest https://www.youtube.com/watch?v=6RRSxWtJPUw

From Extreme to everyday https://www.youtube.com/watch?v=tQ_oo7_36r8

Reference https://nanoporetech.com/how-it-works

Nanopore Sequencing of Ebola Viruses Under Outbreak Conditions https://www.youtube.com/watch?v=SYBzPEoENWI; https://www.nature.com/articles/nature16996

Read length and capacity go beyond



Epigenomics

The *epigenome* is a multitude of chemical compounds that can tell the *genome* what to do.

Histone modifications
 DNA Methylation



Directly detect DNA and RNA methylation with high reproducibility and low bias

Using nanopore sequencing, researchers have directly identified DNA and RNA base modifications at nucleotide resolution, including 5mC, 5hmC, 6mA, and BrdU in DNA, and m6A in RNA, with detection of other natural or synthetic epigenetic modifications possible through training basecalling algorithms.

One of the most widespread genomic modifications is 5-methylcytosine (5mC), which most frequently occurs at CpG dinucleotides. Compared to whole-genome bisulfite sequencing, the traditional method of 5mC detection, nanopore technology calls a higher number of CpG positions in the genome, requires less sequencing data, and shows more even genomic coverage with considerably lower GC bias; analysis runtime is also significantly shorter (Figure 1).

https://nanoporetech.com/applications/investigation/epige netics-and-methylation-analysis



New techniques that transformed genomics in addition to long reads: Chromosome conformation capture (previously used epigenetic dimension of chromosomes)



Lieberman-Aiden (2009) Science 10.1126/science.1181369



Lieberman-Aiden (2009) Science 10.1126/science.1181369

Summary: evolution of sequence reads

DNA fragment (300-600bp)

Paired end reads ; ~150bp sequenced on both ends of a DNA fragment. We know these read pairs belong to the same fragment. Illumina ; high accuracy ; "short" reads Still needed with its high depth and accuracy

DNA fragment (1-30+++ kb)



All these reads belong to the same fragment

10X technology sequenced in Illumina ; high accuracy ; 'linked' reads

Whole read (very long) sequenced!

Oxford Nanopore or Pacbio ; still errorneous; contain modification information

Analysis approaches



OPINION

The real cost of sequencing: higher than you think!

Andrea Sboner^{1,2}, Xinmeng Jasmine Mu¹, Dov Greenbaum^{1,2,3,4,5}, Raymond K Auerbach¹ and Mark B Gerstein^{*1,2,6}



Four situations you are most likely to encounter

Genome reference is available (for example, humans):

- Re-sequence (DNA, RNA)
- Map (align) sequence to the genome

Genome reference is NOT available

Assemble the reads to get the genome

Counting:

- For a given region (gene) we want to know how much.→ gene expression or metagenomics
- Statistics

What is an alignment? (mapping)

Align the following two sequences:

```
ATTGAAAGCTA
GAAATGAAAAGG
1:
--ATTGAAA-GCTA
| | | | | | |
GAAATGAAAAGG--
```

Scoring scheme is needed: 1 for match -1 for mismatch -2 for gap

```
2:
ATTGAAA-GCTA---
|||||||||
```

insertions / deletions (indels) mismatches
Which alignment is better?

Mapping

Reference genome depicting two example genes



■ Discordant reads (structural variant) ▲ Variant base (coding)

Variant base (noncoding)

Variant base (coding)

doi:10.1038/nrgastro.2012.126

Long read able to uncover long SV



https://www.nature.com/articles/s41467-017-01343-4

Assembly



Genome (3.000.000 letters)

Genome (3.000.000 letters)

Long read + HiC to produce a reference assembly (since 2017)



b

| | Goat CHIR_1.0 | Goat ARS1 | Human GRCh38 |
|------------------------------------|------------------|--------------|-----------------|
| Total sequence length | 2.6 Gb | 2.9 Gb | 3.2 Gb |
| Total assembly gap length | 140 Mb | 38 Mb | 160 Mb |
| Gaps between scaffolds | 411 | 0 | 349 |
| Number of scaffolds | 77,431 | 29,907 | 735 |
| Scaffold N50 | 14 Mb | 87 Mb | 67 Mb |
| Number of contigs | 337,494 | 30,399 | 1,385 |
| Contig N50 | 18.9 kb | 26.2 Mb | 56.4 Mb |
| Number of chromosomes and plasmids | 30 | 31 | 25 |





https://www.nature.com/articles/ng.3824

Toward a genome sequence for every animal: Where are we now?

Scott Hotaling^{a,1}, Joanna L. Kelley^a, and Paul B. Frandsen^{b,c,d,1}

Edited by Gene E. Robinson, University of Illinois at Urbana–Champaign, Urbana, IL, and approved October 28, 2021 (received for review August 4, 2021)





Annotation



Nature Reviews | Genetics

Yandell and Ence Nature Genetics Review (2012)



Cook et al (2017) Plant Physiology

Case studies - genomics

Scenarios now and then

- 1. [lab/hospital/mountain/sea] Collect samples (1.1, 1.2, 1.3...)
- 2. [lab/hospital] Extract DNA (2.1, 2.2, 2.3...)
- 3. [lab/hospital/company] Sequencing (3.1, 3.2, 3.3...)
- 4. [lab/company] Analysis
- 5. [lab/hospital] Report
- 1. [lab/hospital/mountain/sea] Collect samples -> report



(diagnostic, cheaper, larger-scale..)



Current and future





The Darwin Tree of Life

Reading the genomes of all life: a new platform for understanding our biodiversity.

The Darwin Tree of Life project aims to sequence the genomes of all 70,000 species of eukaryotic organisms in Britain and Ireland. It is a collaboration between biodiversity, genomics and analysis partners that hopes to transform the way we do biology, conservation and biotechnology.

CREATING A NEW FOUNDATION FOR BIOLOGY

Sequencing Life for the Future of Life

- Sequencing will still be cheaper, read will get longer
- Projects will be bigger



• Standard labs will be able to generate collections of themselves



Classical genetics



Figure 2 | **Strategies for finding disease-causing rare variants using exome sequencing.** Four main strategies are illustrated. **a** | Sequencing and filtering across multiple unrelated, affected individuals (indicated by the three coloured circles). This approach is used to identify novel variants in the same gene (or genes), as indicated by the shaded region that is shared by the three individuals in this example. **b** | Sequencing and filtering among multiple affected individuals from within a pedigree (shaded circles and squares) to identify a gene (or genes) with a novel variant in a shared region of the genome. **c** | Sequencing parent–child trios for identifying *de novo* mutations. **d** | Sampling and comparing the extremes of the distribution (arrows) for a quantitative phenotype. As shown in panel **d**, individuals with rare variants in the same gene (red crosses) are concentrated in one extreme of the distribution.

http://www.nature.com/nrg/journal/v12/n11/pdf/nrg3031.pdf

Comparative genomics / Phylogenomics



Guojie Zhang et al. Science (2014)



Nature Reviews | Genetics

Roger & Gibbs Nature Reviews Genetics (2014)

Comparative genomics

Genomics of the origin and evolution of Citrus

b





Wu et al., Nature (2018)

Population genomics



Novembre et al Nature (2008)



http://www.genomenext.com/casestudies_post/populationscale-analysis-genomic-samples-analyzed-from-2504individuals-in-1-week/

FOCUS ON GENOMES OF ICELANDERS

ARTICLES

genetics

Large-scale whole-genome sequencing of the Icelandic population



A collection of Icelandic genealogical records dating back to the 1700s.

Nature Genetics volume 47, pages 435-444 (2015)

Here we describe the insights gained from sequencing the whole genomes of 2,636 Icelanders to a median depth of 20×.



The blood of a thousand Icelanders. Photo: Chris Lund



RARE GENETIC VARIANTS IN HEALTH AND DISEASE

The project is taking a two-pronged approach to identify rare variants and their effects:

•by studying and comparing the DNA of 4,000 people whose physical characteristics are well documented, the project aims to identify those changes that have no discernible effect and those that may be linked to a particular disease;

•by studying the changes within protein-coding areas of DNA that tell the body how to make proteins of 6,000 people with extreme health problems and comparing them with the first group, it is hoped to find only those changes in DNA that are responsible for the particular health problems observed.

The project received a **£10.5 million** funding award from Wellcome in March 2010 and sequencing started in late 2010. For more information, please use the links on the right hand side.

https://www.uk10k.org/

United Kingdom Genomics England 2012-100,000 Genomes: rare disease, cancer £350M (USD\$485M) Scottish Genomes £6M (USD\$8M) Welsh Genomics for Precision Medicine £6.8M (USD\$9M) Northern Ireland Genomic Medicine Centre £3.3M (USD\$4.6M)

Switzerland Swiss Personalized Health Network 2017-2020 CHF68M (USD69M)

Netherlands

Rare disease

RADICON-NL 2016-2025

Health Research Infrastructure

Japan

cohorts, drug discovery JPY10.2B (USD\$90.05M)

Japan Genomic Medicine Program, 2015-

Infrastructure, clinical and population-based

France Genomic Medicine Plan 2016-2025 Rare disease, cancer, diabetes €670M (USD\$799M)

Estonia Estonian Genome Project 2000 -Infrastructure and population-based cohort 2017: €5M for 100,000 individuals

> Finland National Genome Strategy 2015-2020 Infrastructure €50M (\$USD 59M)

> > Denmark Genome Denmark 2012-DK 86M (USD\$13.5M) FarGen 2011- 2017 DK 10M (USD\$1.6M) Infrastructure, population-based cohort, pathogen project

Turkey

Turkish Genome Project 2017-2023 Infrastructure, clinical and populationbased cohorts

China Precision Medicine Initiative

Australia

Australian Genomics 2016-2021 Infrastructure, rare disease and cancer AUD\$125M (USD\$95M) Genomics Health Futures Mission 2018-2028 AUD\$500M (USD\$372M)

Stark et al (2019) AJHG

United States of America National Human Genome Research Institute 2007-Infrastructure and clinical cohorts **USD\$427M** All of Us 2016-2025 Population cohort USD\$500M (first two years)

Brazil 2015-

Saudi Human Genome Program, 2013-

Infrastructure, population cohort

Qatar Qatar Genome 2015-

累計收案數 統計至2021年08月31日止(請按此) 社區民眾收案數 151,406 參與個案總數 37,508 完成第一輪追蹤個案總數 醫學中心患者收案數 7,387 參與個案總數 1,418 完成第一輪追蹤個案總數 422 完成第二輪追蹤個案總數 98 完成第三輪追蹤個案總數 8 完成第四輪追蹤個案總數

The Cumulative 累計收案數

統計至2019年07月31日止(請按此) 社區民眾收案數 118,548 參與個案總數 24,936 完成第一輪追蹤個案總數

醫學中心患者收案数 3,145 <sup>參與個案總數</sub>
659
完成第一輪追蹤個案總數
104
完成第二輪追蹤個案總數</sup>

The Cumulative 累計收案數

統計至2019年01月31日止(請按此) 社區民眾收案數 109,059 參與個案總數 22,502 完成第一輪追蹤個案總數

醫學中心患者收案數
1,862
參與個案總數
320
完成第一輪追蹤個案總數
8
完成第二輪追蹤個案總數



Precision medicine 精準醫學



Ashley (2016) Nature Review Genetics

Outline of precision medicine



Morash et al (2018) Journal of Personalized Medicine

Summary of outcomes in Oncology PM Studies

| Study | Sample Size | Most Prevalent Tumor Types | Outcomes Reported |
|--|---|--|--|
| Tsimberidou et al. <i>Clin. Cancer Res.</i> 2012 [5] | 291 patients with one molecular aberration (175 treated with matched therapy, 116 control) | Colorectal, melanoma, lung, ovarian | Matched group had improved ORR (27% vs. 5%), TTF (median 5.2 vs. 2.2 month), OS (median 13.4 vs. 9.0 month) |
| Radovich et al. <i>Oncotarget</i> 2016 [6] | 101 patients with sequencing and follow up (44 treated with matched therapy, 57 control) | Soft tissue sarcoma, breast, colorectal | Matched group had improved PFS (86 vs. 49 days) |
| Schwaederle et al. <i>Mol. Cancer Ther.</i> 2016 [7] | 180 patients with sequencing and follow up (87 treated with matched therapy, 93 control) | Gastrointestinal, breast, brain | Matched group had improved PFS (4.0 vs. 3.0 month), TRR (34.5% vs. 16.1% achieving SD/PR/CR) |
| Kris et al. <i>JAMA</i> 2014 [8] | 578 patients with oncogenic driver and followup (260 with matched therapy, 318 control) | Lung only | Matched group had improved survival (median 3.5 vs. 2.4 years) |
| Aisner et al. J. Clin. Oncol. 2016 [9] | 187 patients with targetable alteration and follow up (112 with matched therapy, 74 control) | Lung only | Matched group had improved survival (median 2.8 vs. 1.5 years) |
| Stockley et al. Genome Med. 2016 [10] | 245 patients with sequencing matched to clinical trials (84 on matched trial, 161 control) | Gynecological, lung, breast | Matched group had improved ORR (19% vs. 9%) |
| LeTourneau et al. <i>Lancet</i> Oncol. 2015 [11] | RCT with 195 patients with molecular aberration (99 treated with matched therapy, 96 control) | Gastrointestinal, breast, brain | No difference in PFS between groups |

ORR = overall response rate, TTF = time to treatment failure, OS = overall survival, PFS = progression free survival, TRR = tumor response rate, SD = stable disease, PR = partial response, CR = complete response, RCT = randomized controlled trial. Matched group indicates patients matched to a therapy based on sequencing results.

Morash et al (2018) Journal of Personalized Medicine

Population genomics of pathogens



Mutreja *et al.*, Nature Genetics (2011)

0.02
2013

2016

8000家庭破碎 聯合國遭控傳染霍亂 2013-10-11 by: PMM ●\$725 ● f ● 一直以來,聯合國給世人的印象多是促進世界永續發展的正面印象,但對海地居民來說,聯合國卻成 了當地人最恐懼的創子手,最新報導就指出,因聯合國駐軍而散佈的霍亂已經造成8,000人死亡。 BBC綜合報導,聯合過派駐的維和部隊(UN peacekeepers)意外將細菌帶到海地境內,在當地造成霍亂大流行,自2010年爆發至今,霍亂已經 在海地造成8,000人病死,這也讓海地成為目前全世界霍亂疫病最嚴重的地區。

聯合國是兇手

儘管許多調查指出聯合國就是霍亂源頭,但海地數度請願要求補償未 果,現在海地的代表律師團就上訴紐約法院,控告聯合國是造成海地霍 亂疫情的元凶。

聯合國坦承:我們將霍亂帶進了海地

| 016-08-19 by: 網仔 @1504 | | Ψ. |
|------------------------|--|----|
|------------------------|--|----|

將近六年的時間,聯合國終於承認海地的霍亂疫情與他們有關。到目前為止,已經有數十萬名居民感染上霍亂、一萬名海地人因霍亂而去世。

f 😏

維和部隊惹的禍?

由於海地過去都沒有類似霍亂症狀的疾病,部分專家也發現海地的霍亂 細菌種類與尼泊爾的種類是一樣的,因此懷疑是聯合國在尼泊爾的維和 部隊將霍亂弧菌帶進海地。但將近六年來,聯合國一直都否認這樣的指 控。

聯合國坦承與疫情爆發有關

在本周三(17),聯合國副發言人哈奇(Farhan Haq)聲明:「過去幾年 來,聯合國有鑑於海地初期的瘟疫爆發與我們有些關係,聯合國決定要 多做些什麼。」他也強調聯合國會在接下來兩個月內有所行動。

About

The Project

This project is developing an end-to-end system for processing samples from viral outbreaks to generate real-time epidemiological information that is interpretable and actionable by public health bodies. Fast evolving RNA viruses (such as Ebola, MERS, SARS, influenza etc) continually accumulate changes in their genomes that can be used to reconstruct the epidemiological processes that drive the epidemic. Based around a recently developed, single-molecule portable sequencing instrument, the Oxford Nanopore Technology MinION, we are creating a 'lab-in-a-suitcase' that can be deployed to remote and resource-limited locations. Targeting a wide-range of emerging viral diseases, the sequencing generation will be closely linked to the analysis platform to integrate these data and associated epidemiological knowledge to reveal the processes of transmission, virus evolution and epidemiological linkage with extremely rapid turn-around. This real-time approach will provide actionable epidemiological insights within days of samples being taken from patients.

https://artic.network/ncov-2019

nCoV-2019



There is a pressing need to understand more about the short-term genomic epidemiology and evolution of the recently described novel coronavirus (nCoV-2019). Initial cases were in Wuhan City, Hubei Province, China but now cases have been confirmed both more widely in China and internationally.

Viral genome data generated prospectively during outbreaks can help provide information about relatedness to other viruses, mode and tempo of evolution, geographical spread and adaptation to human hosts. This information can be used to assist in epidemiological investigations, particularly when combined with other types of data (e.g. case counts).

The ARTIC network is making available a set of materials (see below) to assist groups in sequencing the virus including a set of primers, laboratory protocols, bioinformatics tutorials and datasets. These are mainly focused around the use of the portable Oxford Nanopore MinION sequencer, although aspects of the protocol such as the primer scheme and sample amplification may be generalised to other sequencing platforms.

https://artic.network/ncov-2019



HELP DOCS BLOG LOGIN

Nextstrain

Real-time tracking of pathogen evolution

Nextstrain is an open-source project to harness the scientific and public health potential of pathogen genome data. We provide a continually-updated view of publicly available data alongside powerful analytic and visualization tools for use by the community. Our goal is to aid epidemiological understanding and improve outbreak response. If you have any questions, or simply want to say hi, please give us a shout at hello@nextstrain.org.

https://nextstrain.org/



https://nextstrain.org/

다 GISAID Initiative Retweeted

Nextstrain @nextstrain · Mar 6

nextstrain.org/ncov now updated with the first sequence from New Zealand A. The NZ seq groups with other sequences with travel history to Iran (circled), as expected. Sequenced by @ESRNewZealand @MathStorey @Joepdl @sciolato using @NetworkArtic & #RAMPART. Data via @GISAID



Nextstrain @nextstrain · 2h

Thanks to **#opendata** sharing by **@dasmaninstitute @KUWAIT_MOH @FahdAlMulla @KATarinambraun @GageKMoreno @tcflab @dho_lab @ESRNewZealand @MathStorey @Joepdl @sciolato** & **@GISAID nextstrain.org/ncov** is updated with 7 new sequences from Kuwait, Wisconsin, & New Zealand!

#COVID19



https://nextstrain.org/narratives/ncov/sit-rep/en/2020-03-20



Yesterday, the Public Health Wales Specialist virology Centre passed the first two positive samples in Wales to the team in the Pathogen Genomics Unit to perform whole genome sequencing

Q 1 1 2

♡ 13



Catherine Moore 🦠 🖉 🔤 🚟 👬 @SmallRedOne · Mar 7 Today... V

,↑,





Catherine Moore Catherine Moore Catherine Moore Mar 7 @tomrconnor tells me that our sequences are almost ready for public release through **#GISAID** for addition to the global dataset. This is incredible.

Catherine Moore 🦠 🖉 🔜 🖀 🗮 @SmallRedOne - Mar 7 And we're live





https://www.science.org/news/2021/08/new-sars-cov-2-variants-havechanged-pandemic-what-will-virus-do-next https://www.science.org/doi/epdf/10.1126/science.acx8919

20 August 2021

Human gut microbiome



Human gut microbiome

Vol 464 4 March 2010 doi:10.1038/nature08821

ARTICLES

A human gut microbial gene catalogue established by metagenomic sequencing

Junjie Qin¹*, Ruiqiang Li¹*, Jeroen Raes^{2,3}, Manimozhiyan Arumugam², Kristoffer Solvsten Burgdorf⁴, Chaysavanh Manichanh⁵, Trine Nielsen⁴, Nicolas Pons⁶, Florence Levenez⁶, Takuji Yamada², Daniel R. Mende², Junhua Li^{1,7}, Junming Xu¹, Shaochuan Li¹, Dongfang Li^{1,8}, Jianjun Cao¹, Bo Wang¹, Huiqing Liang¹, Huisong Zheng¹, Yinlong Xie^{1,7}, Julien Tap⁶, Patricia Lepage⁶, Marcelo Bertalan⁹, Jean-Michel Batto⁶, Torben Hansen⁴, Denis Le Paslier¹⁰, Allan Linneberg¹¹, H. Bjørn Nielsen⁹, Eric Pelletier¹⁰, Pierre Renault⁶, Thomas Sicheritz-Ponten⁹, Keith Turner¹², Hongmei Zhu¹, Chang Yu¹, Shengting Li¹, Min Jian¹, Yan Zhou¹, Yingrui Li¹, Xiuqing Zhang¹, Songgang Li¹, Nan Qin¹, Huanming Yang¹, Jian Wang¹, Søren Brunak⁹, Joel Doré⁶, Francisco Guarner⁵, Karsten Kristiansen¹³, Oluf Pedersen^{4,14}, Julian Parkhill¹², Jean Weissenbach¹⁰, MetaHIT Consortium[†], Peer Bork², S. Dusko Ehrlich⁶ & Jun Wang^{1,13}

nature

doi:10.1038/nature08821

Human gut microbiome



We can check which OTUs constitute the clustering (and separation) patterns

- -> Biology
- -> Biomarkers

Tracking microbiome on a daily scale



David et al. Genome Biology 2014, 15:R89

Tracking microbiome spanning 6 years



Species-Level Deconvolution of Metagenome Assemblies with Hi-C–Based Contact Probability Maps

Joshua N. Burton,¹ Ivan Liachko,¹ Maitreya J. Dunham,² and Jay Shendure² Department of Genome Sciences, University of Washington, Seattle, Washington 98195-5065





| Table 2 | Sequencing | libraries | used in | MetaPhase | analyses |
|---------|------------|-----------|---------|-----------|----------|
|---------|------------|-----------|---------|-----------|----------|

| Sample | Library Type | Read Length, bp | Read Pairs, millions |
|--------|--------------|-----------------|----------------------|
| M-Y | Shotgun | 101 | 85.7 |
| | Mate-pair | 100 | 9.2 |
| | Hi-C | 100 | 81.0 |



Other fields transformed by genomics







ancient DNA



ORIGINAL ARTICLES Green, R. E. et al. A draft sequence of the Neandertal genome. *Science* **328**, 710–722 (2010) | Meyer, M. et al. A high-coverage genome sequence from an archaic Denisovan individual. *Science* **338**, 222–226 (2012) | Rasmussen, M. et al. Ancient human genome sequence of an extinct Palaeo-Eskimo. *Nature* **463**, 757–762 (2010)

FURTHER READING Slon, V. et al. The genome of the offspring of a Neanderthal mother and a Denisovan father. *Nature* **561**, 113–116 (2018) | Haak, W. et al. Massive migration from the steppe was a source for Indo-European languages in Europe. *Nature* **522**, 207–211 (2015) | Sankararaman, S. et al. The date of interbreeding between Neandertals and modern humans. *PLoS Genet.* **8**, e1002947 (2012) "

Ancient DNA research has been limited only by the technology, and never by a lack of interesting questions to be asked





Genomic sequencing

- 1. The Human Genome Project
- 2. Sequencing the unculturable majority
- 3. Sequencing the next generation
- 4. ChIP-seq captures the chromatin landscape
- 5. The dawn of personal genomes
- 6. A sequencing revolution in cancer
- 7. Transcriptomes a new layer of complexity
- 8. Long reads become a reality
- 9. Exploring whole exomes
- 10. Probing nuclear architecture with Hi-C
- 11. Sequencing one cell at a time
- 12. Waking the dead: sequencing archaic hominin genomes
- 13. Cataloguing a public genome
- 14. Our most elemental encyclopaedia
- 15. Pan-genomes: moving beyond the reference
- 16. Genomes go platinum
- 17. Filling in the gaps telomere to telomere

Break here

Transcriptomics / RNAseq

Types of RNA





RIO • ARES • HANNON • NILSEN

http://jura.wi.mit.edu/bio/education/hot_topics/RNAseq/RNA_Seq.pdf

Gene and isoforms



Conesa et al. Genome Biology (2016) 17:13 DOI 10.1186/s13059-016-0881-8

Genome Biology

REVIEW

Open Access

A survey of best practices for RNA-seq data analysis

Ana Conesa^{1,2*}, Pedro Madrigal^{3,4*}, Sonia Tarazona^{2,5}, David Gomez-Cabrero^{6,7,8,9}, Alejandra Cervera¹⁰, Andrew McPherson¹¹, Michał Wojciech Szcześniak¹², Daniel J. Gaffney³, Laura L. Elo¹³, Xuegong Zhang^{14,15} and Ali Mortazavi^{16,17*}

RNA-seq data generation



Downstream analysis

Griffith et al (2015) Plos Computational Biology

RNAseq analysis workflow for differential expression (generalized)



Stark, Grzelak and Hadfield (2019) Nature Reviews Genetics

REVIEWS

RNA sequencing: the teenage years

Rory Stark¹, Marta Grzelak¹ and James Hadfield^{2*}

Abstract | Over the past decade, RNA sequencing (RNA-seq) has become an indispensable tool for transcriptome-wide analysis of differential gene expression and differential splicing of mRNAs. However, as next-generation sequencing technologies have developed, so too has RNA-seq. Now, RNA-seq methods are available for studying many different aspects of RNA biology, including single-cell gene expression, translation (the translatome) and RNA structure (the structurome). Exciting new applications are being explored, such as spatial transcriptomics (spatialomics). Together with new long-read and direct RNA-seq technologies and better computational tools for data analysis, innovations in RNA-seq are contributing to a fuller understanding of RNA biology, from questions such as when and where transcription occurs to the folding and intermolecular interactions that govern RNA function.

Stark, Grzelak and Hadfield (2019) Nature Reviews Genetics

Evolution of RNAseq over time (from SRA)



Berge et al., Annual Review of Biomedical Data Science (2019)

Further advances Single cell RNAseq (ScRNAseq)

single cell sequencing



ORIGINAL ARTICLE Tang, F. et al. mRNA-Seq whole-transcriptome analysis of a single cell. *Nat. Methods* **6**, 377–382 (2009)

FURTHER READING Islam, S. et al. Characterization of the single-cell transcriptional landscape by highly multiplex RNA-seq. *Genome Res.* **21**, 1160–1167 (2011) | Ramsköld, D. et al. Full-length mRNA-Seq from single-cell levels of RNA and individual circulating tumor cells. *Nat. Biotechnol.* **30**, 777–782 (2012) | Macosko, E. Z. et al. Highly parallel genome-wide expression profiling of individual cells using nanoliter droplets. *Cell* **161**, 1202–1214 (2015) | Klein, A. M. et al. Droplet barcoding for single-cell transcriptomics applied to embryonic stem cells. *Cell* **161**, 1187–1201 (2015) | Haber, A. L. et al. A singlecell survey of the small intestinal epithelium. *Nature* **551**, 333–339 (2017) | Vento-Tormo, R. et al. Single-cell reconstruction of the early maternal-fetal interface in humans. *Nature* **563**, 347–353 (2018) single-cell techniques [have] enabled analysis of cell states, ... diseased states and tumour heterogeneity

Evolution of single-cell isolation



Microfluidic isolation in reagent- filled droplets



Huang et al., Experimental & Molecular Medicine (2018) 50:96

Structure of the barcode primer bead



Spatialomics

1. Spatial encoding requires a frozen tissue section to be applied to oligo- arrayed microarray slides or to 'pucks' of densely packed oligo- coated beads.



2. The mRNA diffuses to the slide surface and hybridizes to oligo- dT cDNA synthesis primers that encode UMIs and spatial barcodes. It is then reverse transcribed to produce cDNA, which is pooled for library preparation and sequencing.

3. Computational analysis of the spatialomics data maps sequence reads back to their spatial coordinates after DGE analysis and allows differential spatial expression to be visualized.

Stark *et al.*, Nature Review Genetics (2019)

Applications of scRNAseq computational approaches 1. Cell type identification



Applications of scRNAseq computational approaches 2. Cell hierarchy reconstruction



Applications of scRNAseq computational approaches 2. Cell hierarchy reconstruction



Applications of scRNAseq computational approaches 3. Inferring regulatory networks

Gene transcription "on"



Identifying modules of co-regulated genes

Applications of scRNAseq computational approaches 3. Inferring regulatory networks


Applications of scRNAseq computational approaches 3. Inferring regulatory networks



Huang et al., Experimental & Molecular Medicine (2018) 50:96

Other approaches

nascent RNA

Essentially enrich newly transcribed RNAs in a cell and compare to control (mature RNA)



Stark et al., Nature Review Genetics (2019)

translatome

- RNA-sequencing from ribsomally bound RNA
- mRNA ribosome density correlates with the protein synthesis level



Stark et al., Nature Review Genetics (2019)

RNA-RNA interaction RNA-protein interaction

- A) Probe structured (ddRNA) or unstructured (ssRNA) RNA in transcriptome level
- B) Crosslinking interacting RNA with biotinylated psoralen
- C) Crosslinking immunoprecipitation of RNA followed by sequencing



Stark et al., Nature Review Genetics (2019)

Single cell sequencing + multi omic perspective

Overview of current methods for single cell data integration



Example of experimental methods for performing single-cell multimodal measurements



Multi-modal data can lead to better power at identifying cell states



Spatial omics + scRNA-seq



Spatial omics + scRNA-seq



Mapping smFISH cells onto scRNA-seq data allows the transfer of cell-type classifications derived from transcriptome-wide gene expression measurements to be transferred to the spatially resolved cells

Type cells in situ

Stuart and Satija (2019) Nature Review Genetics

mapping scRNA-seq data onto smFISH-profiled spatial coordinates can allow scRNA-seq data from dissociated cells to be placed back into their spatial context

Map cells to spatial coordinates

Spatial omics + scRNA-seq



By mapping scRNA-seq-profiled cells onto spatially resolved coordinates through the integration with smFISH data, **spatial patterns of gene expression can be predicted for any gene measured in the scRNA-seq data set**. Through these predictions, novel spatial patterns of gene expression may be identified through the analysis of genes that were not profiled by smFISH

Stuart and Satija (2019) Nature Review Genetics

X

a multi-omic perspective

Multiple omics data types



- Genome first or Phenotype first or environment first?
- Genome first -> GWAS
- "Locus-centered integration of additional omics layers can help to identify causal single nucleotide polymorphisms(SNPs) and genes at GWAS loci and then to examine how these perturb pathways leading to disease"

Hain et al (2017) Genome Biology

Integrating multi-omics to network

Various additional data can then be used to enrich and extract biological relevant information from the network



Dam et al (2016) Briefings in Bioinformatics



"a paradigm for future multi-omic studies of the human microbiome"



Proctor, L.M., Creasy, H.H., Fettweis, J.M. *et al.* The Integrative Human Microbiome Project. *Nature* **569**, 641–648 (2019). https://doi.org/10.1038/s41586-019-1238-8

Summaries

VIEWPOINT

The road ahead in genetics and genomics

Amy L. McGuire, Stacey Gabriel, Sarah A. Tishkoff, Ambroise Wonkam, Aravinda Chakravarti, Eileen E. M. Furlong, Barbara Treutlein, Alexander Meissner, Howard Y. Chang, Núria López-Bigas, Eran Segal and Jin-Soo Kim

Abstract | In celebration of the 20th anniversary of *Nature Reviews Genetics*, we asked 12 leading researchers to reflect on the key challenges and opportunities faced by the field of genetics and genomics. Keeping their particular research area in mind, they take stock of the current state of play and emphasize the work that remains to be done over the next few years so that, ultimately, the benefits of genetic and genomic research can be felt by everyone.

- 1. Making genomics truly equitable
- 2. Genome sequencing at population scale
- 3. A global view of human evolution
- 4. African genomics the next frontier
- 5. Decoding multifactorial phenotypes
- 6. Enhancers and embryonic development
- 7. Spatial multi- omics in single cells
- 8. Unravelling the layers of the epigenome
- 9. Long non- coding RNAs: a time to build
- 10. FAIR genomics to track tumorigenesis
- 11. Integrating genomics into medicine
- 12. CRISPR genome editing enters the clinic

New challenges

- So much data
- Technology advancement
- Integrating different kinds of data (multi-omic)
- High performance
- Reproducibility crisis
- Bioinformaticians as a profession
- Only biology has a specific term to refer to the use of computers in this discipline ('bioinformatics')
- Proper integration into academic curriculums

A personal take on science and society

World view

Biology must generate ideas as well as data



By Paul Nurse

It would have been a pity if Darwin had stopped thinking after describing the shapes and sizes of finch beaks."

Accepting a Nobel prize nearly two decades ago, my old friend Sydney Brenner had a warning for biology. **"We are drowning in a sea of data and starving for knowledge,"** he said. That admonishment, from one of the founders of molecular biology, who established the nematode worm *Caenorhabditis elegans* as a model organism, is even more relevant to biology today.

13 September 2021

https://www.nature.com/articles/d41586-021-02480-z

Yanai and Lercher *Genome Biology* (2020) 21:231 https://doi.org/10.1186/s13059-020-02133-w

Genome Biology

EDITORIAL

A hypothesis is a liability

Itai Yanai^{1*} and Martin Lercher^{2*}

" 'When someone seeks,' said Siddhartha, 'then it easily happens that his eyes see only the thing that he seeks, and he is able to find nothing, to take in nothing. [...] Seeking means: having a goal. But finding means: being free, being open, having no goal.' " Hermann Hesse



Open Access

Shift in paradigm 2005-2021 (My personal take)

- Genome and transcriptome sequencing projects are
 - being done on a per-lab basis and no longer exclusive to sequencing centers
 - moving away from exploration to question orientated.
- Data being produced on a much faster speed at a much higher throughput, and a much cheaper scale
- More methods, analysis, tools, experiments...
 - Not always better

It is an exciting time to be in

Thank you